Marta Bałajewicz-Nowak, MD, PhD Mail: marta.balajewicz@gmail.com



The gonads do not acquire male or female morphological characteristics until the **seventh week of development.**

Gonads appears initially as a pair of longitudinal ridges – genital/gonadal ridges.

Primordial germ cells (they're formed in the epiblast during the second week and than move to the wall of the yolk sac) invade the genital ridges – 6th week.

PGCs have inductive influence on development of gonads.

Epithelial cells proliferate and penetrate the underlying mesenchyme – form primitive sex cords = indifferent gonad



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Source: Molina PE: Endocrine Physiology, 3rd Edition: http://www.accessmedicine.com



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Source: Mescher AL: Junqueira's Basic Histology: Text and Atlas, 12th Edition: http://www.accessmedicine.com



SERTTOLI CELLS •Support, protection, and nutrition of the developing spermatogenic cells spermatocytes, spermatids, and sperm are isolated from plasma proteins and nutrients

•Exocrine and endocrine secretion. a fluid used for sperm transport in the direction of the genital ducts.

•Phagocytosis. During spermiogenesis, excess cytoplasm shed as residual bodies is phagocytosed and digested by Sertoli cell lysosomes. No proteins from sperm normally pass back across the <u>blood-testis barrier.</u>

LEYDIG CELLS

by the blood-testis barrier,

Leydig cells are responsible for the production of 95% of adult male testosterone.

Source: Molina PE: Endocrine Physiology, 3rd Edition: http://www.accessmedicine.com





12th Edition: http://www.accessmedicine.com

WHO laboratory manual for the Examination and processing of human semen

FIFTH EDITION

Table A1.1 Lower reference limits (5th centiles and their 95% confidence intervals) for semen characteristics

Parameter		Lower reference limit
Semen volume (ml)		1.5 (1.4–1.7)
Total sperm number (10 ⁶ per ejaculate)		39 (33–46)
Sperm concentration (10 ⁶ per ml)		15 (12–16)
Total motility (PR + NP, %)		40 (38–42)
Progressive motility (PR, %)		32 (31–34)
Vitality (live spermatozoa, %)		58 (55–63)
Sperm morphology (normal forms, %)	$\mathbf{\Lambda}$	4 (3.0–4.0)
Other consensus threshold values		
pH		≥7.2
Peroxidase-positive leukocytes (10 ⁶ per ml)		<1.0
MAR test (motile spermatozoa with bound particles, %)		<50
Immunobead test (motile spermatozoa with bound beads, %)		<50
Seminal zinc (µmol/ejaculate)		≥2.4
Seminal fructose (µmol/ejaculate)		≥13
Seminal neutral glucosidase (mU/ejaculate)		≥20

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Total motility (PR + NP, %)	40 (38–42)
Progressive motility (PR, %)	32 (31–34)
Vitality (live spermatozoa, %)	58 (55–63)
Sperm morphology (normal forms, %)	4 (3.0–4.0)
Other consensus threshold values	
рН	≥7.2
Peroxidase-positive leukocytes (106 per ml)	<1.0
MAR test (motile spermatozoa with bound particles, %)	<50
Immunobead test (motile spermatozoa with bound beads, %)	<50
Seminal zinc (µmol/ejaculate)	≥2.4
Seminal fructose (µmol/ejaculate)	≥13
Seminal neutral glucosidase (mU/ejaculate)	≥20

Table A1.3 Nomenclature related to semen quality

aspermia	no semen (no or retrograde ejaculation)
asthenozoospermia	percentage of progressively motile (PR) spermatozoa below the lower reference limit
asthenoteratozoospermia	percentages of both progressively motile (PR) and morphologically normal spermatozoa below the lower reference limits
azoospermia	no spermatozoa in the ejaculate (given as the limit of quantification for the assessment method employed)
cryptozoospermia	spermatozoa absent from fresh preparations but observed in a centri- fuged pellet
haemospermia (haematospermia)	presence of erythrocytes in the ejaculate
leukospermia (leukocyto- spermia, pyospermia)	presence of leukocytes in the ejaculate above the threshold value
necrozoospermia	low percentage of live, and high percentage of immotile, spermatozoa in the ejaculate
normozoospermia	total number (or concentration, depending on outcome reported)* of spermatozoa, and percentages of progressively motile (PR) and mor- phologically normal spermatozoa, equal to or above the lower reference limits
oligoasthenozoospermia	total number (or concentration, depending on outcome reported)* of spermatozoa, and percentage of progressively motile (PR) spermato- zoa, below the lower reference limits
oligoasthenoterato- zoospermia	total number (or concentration, depending on outcome reported)* of spermatozoa, and percentages of both progressively motile (PR) and morphologically normal spermatozoa, below the lower reference limits
oligoteratozoospermia	total number (or concentration, depending on outcome reported)* of spermatozoa, and percentage of morphologically normal spermatozoa, below the lower reference limits
oligozoospermia	total number (or concentration, depending on outcome reported)* of spermatozoa below the lower reference limit
teratozoospermia	percentage of morphologically normal spermatozoa below the lower reference limit

*Preference should always be given to total number, as this parameter takes precedence over concentration.





Figure 3.31. Female pelvic viscera. A. In this dissection of the female genital organs, the bladder and adjacent anterior pelvis (superior ramus and bodies of pubic bones) have been coronally sectioned and the anterior segment has been removed. On the right side, the uterine tube, ovary, broad ligament, and peritoneum covering the lateral wall of the pelvis have been removed to display the ureter and branches of the internal iliac artery. **B.** This dissection reveals the uterus, ovaries, uterine tubes, and related structures. The broad ligament is removed on the right side.



Source: Cunningham FG, Leveno KJ, Bloom SL, Hauth JC, Rouse DJ, Spong CY: Williams Obstetrics, 23rd Edition: http://www.accessmedicine.com



Source: Schorge JO, Schaffer JI, Halvorson LM, Hoffman BL, Bradshaw KD, Cunningham FG: Williams Gynecology: http://www.accessmedicine.com

Numbers

- In 5th month the number of germ cells reach max. 7 mln.
- The maximal number of oogonia is achieved at the 20th week of gestation, at which time six to seven million oogonia are present in the ovary
- Approximately one to two million oogonia are present at birth with less than 400,000 present at the initiation of puberty, of which less than 500 are destined to ovulate
- Therefore, most female germ cells are lost through atresia



BRAIN CONTROL

What's the role of hypothalamus?

ULTRA SHORT FEEDBACK LOOP

ARCUATE NUCLEUS (Gonadotropin Releasing Hormone)



Hypothalamic-releasing and Hypothalamic-inhibiting hormones

GnRH

- Pulsatile release of gonadotropin-releasing hormone (GnRH) from the hypothalamus stimulates pulsatile pituitary release of luteinizing hormone (LH) and follicle-stimulating hormone (FSH).
- GnRH secretion is regulated by dopamine, serotonin, endorphin, and norepinephrine.
- Hypothalamus is responsive to light, stress, steroids, glucose level, autonimic outputs...



Proliferating/Follicular Phase



Source: Schorge JO, Schaffer JI, Halvorson LM, Hoffman BL, Bradshaw KD, Cunningham FG: Williams Gynecology: http://www.accessmedicine.com

- FSH is responsible for follicular recruitment and growth
- SELECTION WINDOW FSH
 rise leading to the
 development of follicles: a group of antral follicles a cohort begins a phase of semisynchronous growth

Preovulatory Follicle





Source: Cunningham FG, Leveno KJ, Bloom SL, Hauth JC, Rouse DJ, Spong CY: Williams Obstetrics, 23rd Edition: http://www.accessmedicine.com

Ovulation requirements

- Estradiol 仓仓仓
- Progesteron ①
- LH surge 仓仓仓仓

•LH is responsible for ovulation ("trigger") and corpus luteum formation

LH

Collagenase activity Prostaglandin level Metalloproteinase level

Preovulatory Follicle





```
!!!! LH surge
```

Preovulatory growth phase = Meiosis I is completed forming two daughter cells unequal sizes – 23 chromosomes each: SECONDARY OOCYTE FIRST POLAR BODY(with no cytoplasm) – lies between zona pellucida and cell membrane of oocyte in PERIVITELLINE SPACE.

CELLS enters meiosis II and stops in METAPHASE approx. 3 h before ovulation until FERTILIZATION!

Luteal/Secretory Phase

The basement membrane separating the granulosa-lutein and theca-lutein cells break-down and by day 2 postovulation blood vessels and capillaries invade granulosa cell layer. These cells undergo hypertrophy and increase their capacity to synthetize hormones.





- Secondary oocyte (in arrested metaphase of the
 - 2nd meiosis)
- 2 Corona radiata
 - Follicle fluid

3

4

Scattered groups of cumulus cells

*



- 1 Mucus fibers 2
 - (strongly meshed)
- 3 Crypt of a cervix gland



- Mucus fibers 4 (loosely meshed) Portio entrance 5
- The trip from cervix to oviduct requires a minimum of 2 to 7 hours.

Capacitation is what one calls the changes that lead to hyperactivity of the spermatozoon and which later allow the spermatozoon to go through the acrosome reaction. In the human it lasts approx. 7 hours.





- **1** Pores
- 2 Emerging of the acrosomal contents
- 3 Inner acrosomal membrane
- 4 Acrosomal content (enzyme)
- **5** Outer acrosomal membrane
- 6 Cell membrane
- A Head
- **B** Neck
- **C** Mid-piece

Passage through CORONA RADIATA ACROSOME REACTION

Source: Molina PE: Endocrine Physiology, 3rd Edition: http://www.accessmedicine.com



- Post-acrosomal region
- Oolemma with microvilli
- Perivitelline space
- Pellucid zone



FUSION



i Cortical vesicle at the surface of the oocyte



1 2	1rst polar body Nucleus (slightly unpacked) of the
	spermatozoon
3	Proximal centrosome
	of the
л	spermatozoon
4	2nd polar body
	(being formed)
5	Remainder of the
•	mitotic spindle
	with maternal
	chromosomes 1n,1C

The docking triggers a cascade of events with the following goals:

Polyspermy block: The penetration of further sperm cells should be hindered

Hardening of the pellucid zone as a mechanical protection of the embryo

Entry of the spermatozoon into the oocyte Termination of the 2nd meiosis of the oocyte with expulsion of the 2nd polar body

Preparation at the molecular level of the oocyte for unpacking the paternal DNA

4 hours after impregnation



1 Paternal 2 pronucleus 3 Maternal pronucleus Centrosome brought in by the spermatozoon

18 hours after impregnation



Paternal pronucleus Maternal pronucleus Duplicated paternal centrosome "Inner bodies"

1

2

3

4

22hours after impregnations



1

Nucleic membranes of the pronuclei, as they are dissolving Microtubules of 2 the mitotic spindle



Source: Cunningham FG, Leveno KJ, Bloom SL, Hauth JC, Rouse DJ, Spong CY: Williams Obstetrics, 23rd Edition: http://www.accessmedicine.com

- a **diploid** set of chromosomes
- The zygote by definition the first cell of the embryo
- 12-32 cells the morula
- The **blastocyst** when blastocystic cavity appears.



L. morus, mulberry



Source: Cunningham FG, Leveno KJ, Bloom SL, Hauth JC, Rouse DJ, Spong CY: Williams Obstetrics, 23rd Edition: http://www.accessmedicine.com

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The two-cell stage is reached about 30 hours after fertilization.

The four-cell – 40 hours.

12-16 cell stage – 3 days.

Late morula stage – 4 days.

Zona Pellucida disappears at the end of 4th day.

Enters the cavity at the 4th day, when the morula become blastocyst.

Pellucid zone **Trophoblast** (outer cell mass)

1

2

3

4

5

- **Hypoblast** (part of the inner cell mass)
- Blastocyst cavity
- **Epiblast** (part of the inner cell mass)

Blastocyst



Implantation: 6th –7th day

Implantation: 7th –8th day





- Epithelium of the uterine endometrium 1
- 2 Hypoblast
- Syncytiotrophoblast (ST) 3
- 4 Cytotrophoblast (CT)
- Epiblast 5
- 6 Blastocyst cavity

Implantation: 8th day



Implantation: 9th – 10th day



- 1 Syncytiotrophoblast (ST)
- 2 Cytotrophoblast (CT)
- 3 Epiblast
- 4 Hypoblast
- 5 Blastocyst cavity
- 6 Maternal blood capillary
- 7 Amniotic cavity

- 8 Amnioblasts
- 9 Fibrin (closing) plug
- 10 Syncytiothrophoblast lacuna
- **11** Multiplying hypoblast

Implantation: 11th day



- 1 Extra-embryonic mesoderm
- 2 Extra-embryonic reticulum
- **3** Primitive yolk sac (Exocoelomic cavity)
- 4 Cytotrophoblast

Implantation: 12th day



- 5 Lacunae in the reticulum
- 6 Hypoblast
- 7 Heuser's membrane between hypoblast and mesoderm cells

MATERNAL BLOOD ENTERS THE LACUNES.